

Communication

A Short and Highly Stereoselective Synthesis of Cerebrosterol<sup>†</sup>ZHANG, Dong-Hui<sup>a</sup>(张冬辉) ZHOU, Xiang-Dong<sup>b</sup>(周向东) ZHOU, Wei-Shan<sup>\* , a</sup>(周维善)<sup>a</sup> Shanghai Institute of Organic Chemistry, Chinese Academy of Sciences, Shanghai 200032, China<sup>b</sup> Department of Chemistry, The Third Military Medical University, Chongqing 400038, China

The title compound, cerebrosterol, has been synthesized in five steps with 61.5% overall yield and >99.9% *de* from 3 $\alpha$ , 6 $\alpha$ -dimethoxymethoxy-5 $\beta$ -cholane-24-al as a key intermediate, which was prepared from methyl 3 $\alpha$ , 6 $\alpha$ -dihydroxy hyodeoxy-cholanate

**Keywords** cerebrosterol, methyl 3 $\alpha$ , 6 $\alpha$ -dihydroxy hyodeoxy-cholanate, diisopropylzinc, stereoselective synthesis

Cerebrosterol (24*S*-hydroxycholesterol, **1**) formed in small amounts in human and animal brain<sup>1</sup> from cholesterol is important for cholesterol homeostasis in this organ. The excess cholesterol is converted into 24*S*-hydroxycholesterol by a unique brain-specific 24*S*-hydroxylase, which could be readily secreted from the central nervous system into the plasma and be taken up by the liver and further metabolized.<sup>2</sup> Alzheimer's disease (AD) and vascular demented patients appear to have higher circulating levels of cerebrosterol. It is speculated that cerebrosterol may potentially be used as a peripheral indicator of neuronal degeneration and potential state marker for AD.<sup>3</sup> Moreover, cerebrosterol can significantly activate LXR $\alpha$  and LXR $\beta$ , two orphan members of the nuclear receptor superfamily, and play a critical role in the regulation of cholesterol homeostasis. The activated LXRs form a permissive heterodimer with the 9-*cis*-retinoic acid receptor RXR and then regulate the transcriptional expression of their target genes, such as CYP7A,<sup>4</sup> SREBP-1c,<sup>5</sup> ABCB1,<sup>6a,6b</sup> ABCG1,<sup>6c</sup> ABCG5/ABCG8,<sup>6d</sup> ApoE,<sup>7</sup> CETP,<sup>8</sup> LPL<sup>9</sup>, etc. Thus, the cerebrosterol may be a potential drug for treatment for atherosclerosis.

The early synthesis of cerebrosterol relied on resolution via either recrystallization or chromatography of a mixture of 24*R*- and 24*S*-hydroxycholesterol benzoate.<sup>10</sup> Recently, several examples were reported of the stereoselective introduction of 24*S*-hydroxy group into the side chain of cholesterol.<sup>11</sup> Most of them took long steps to construct the side chain of cerebrosterol by using stigmasterol<sup>11a,11b</sup> as starting material, or cholest-25-en-24-one<sup>11c</sup> and desmosterol<sup>11e,11f</sup> as the intermediates. Although in 1995, Okamoto *et al.*<sup>11d</sup> utilized steroidal 3-silylated 24-aldehyde obtained from 3 $\beta$ -hydroxy- $\Delta^5$ -cholonic acid to react with diisopropylzinc using (1*R*, 2*S*)-(+)2-(*N*, *N*-di-*n*-butylamino)-1-phenylpropan-1-ol [(+)-DBNE] as a chiral ligand to give 3 $\beta$ -acetoxycerebrosterol in 63% yield with 87% *de*, the result is not very satisfactory. Furthermore, to prepare 24-aldehyde from 3 $\beta$ -hydroxy- $\Delta^5$ -cholonic acid, which was obtained from 3 $\beta$ -hydroxy- $\Delta^5$ -pregnene-20-one, required lots of steps.<sup>12</sup> Since methyl hyodeoxycholanate (Me-HDCA, **2**) is very cheap and readily available in China and can be efficiently converted to steroidal 3 $\alpha$ , 6 $\alpha$ -bismethoxymethyl ether 24-aldehyde (**3**),<sup>11f</sup> it is used to directly construct the side chain of cerebrosterol via the asymmetric isopropylation of **3**. Moreover, the two retained alkoxy groups at the C-3 and C-6 positions in steroidal 24-aldehyde **3** may facilitate the addition of diisopropylzinc because it was reported that 1 $\alpha$ , 3 $\beta$ -bisilylated cholonic aldehyde reacted with diisopropylzinc catalyzed by chiral  $\beta$ -amino alcohol ligand (+)-DBNE to afford the isopropylated adduct in a much higher yield than 3 $\beta$ -silylated cholonic aldehyde.<sup>11d</sup> More-

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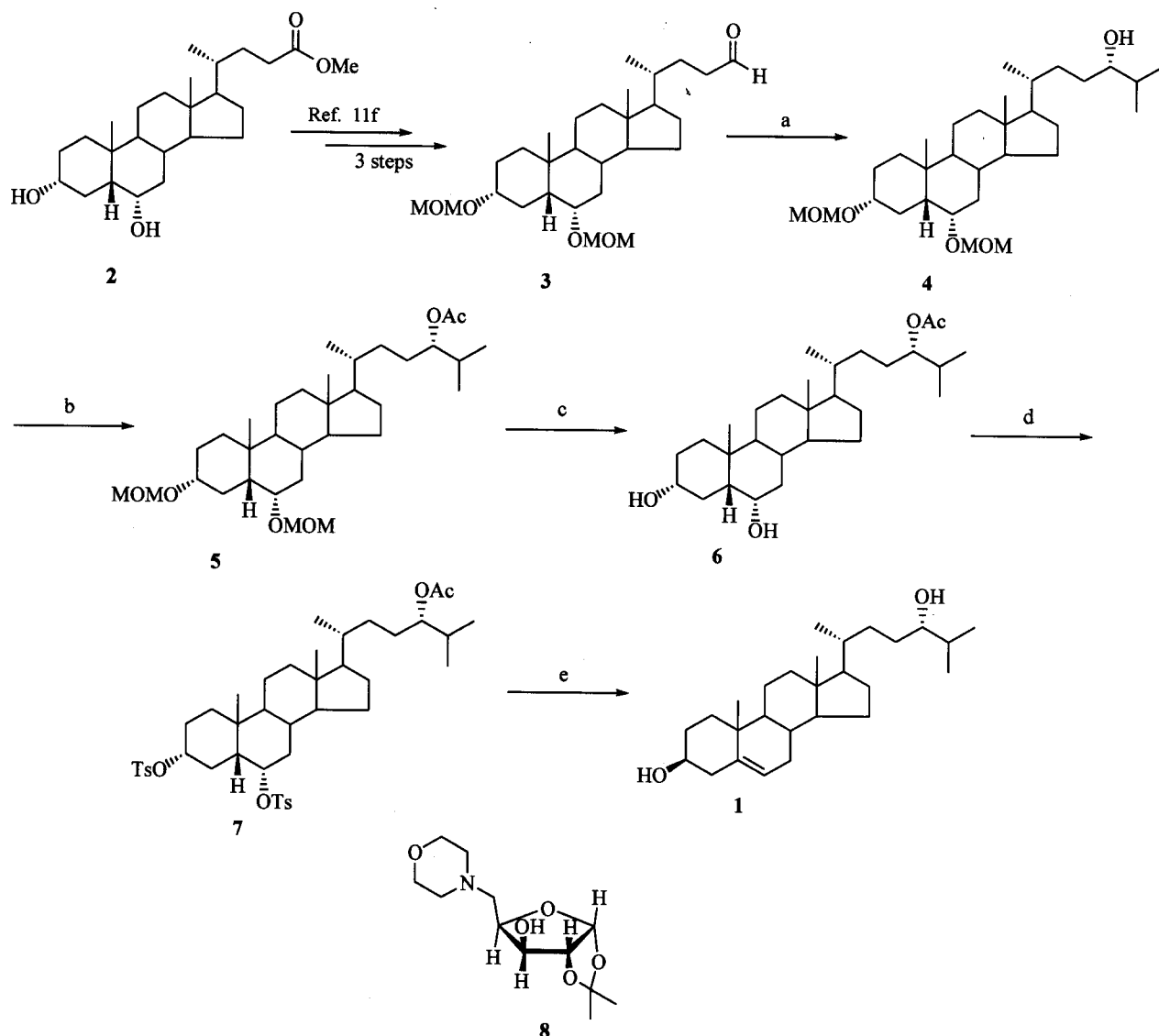
<sup>†</sup>Dedicated to Professor HUANG Yao-Zeng on the occasion of his 90th birthday.

over, the  $3\alpha, 6\alpha$ -bismethoxymethyl ether group in the steroidal 24-aldehyde **3** can be easily converted to the  $\Delta^5$ - $3\beta$ -ol moiety<sup>13</sup> after it is isopropylated. So we select Me-HDCA as the starting material to synthesize cerebrosterol (**1**).

The practically synthetic route to cerebrosterol is outlined in Scheme 1. The key intermediate, steroidal  $3\alpha, 6\alpha$ -bismethoxymethyl ether 24-aldehyde (**3**), was prepared efficiently from Me-HDCA in 85.5% overall yield for three steps according to our previous work.<sup>11f</sup> With the steroidal aldehyde **3** in hand, our attention was

turned to asymmetric isopropylation of the steroidal 24-aldehyde. Very recently, Yang and Cho<sup>14</sup> reported that the enantioselective addition of diisopropylzinc to alkyl and aryl aldehyde catalyzed by a  $\gamma$ -dialkylamino alcohol, 1, 2-*O*-isopropylidene-5-deoxy-5-morprolinó- $\alpha$ -*D*-xylofuranose (**8**), which is easily available from  $\alpha$ -*D*-xylose, with very high enantiomeric excess and yield. We applied this novel ligand for the asymmetric addition of the steroidal aldehyde **3** with diisopropylzinc. Thus **3** was isopropylated with 20 mol% chiral ligand **8** in toluene at 0 °C for 4 h. As expected, the reaction proceeded smoothly

Scheme 1



**Reagents and conditions:** a)  $i$ Pr<sub>2</sub>Zn, ligand **8**, toluene, 0 °C, 85%, *de* > 99.9%; b) Ac<sub>2</sub>O, Py, 99%; c) PPTs, *t*-BuOH, reflux, 88%; d) TsCl, Py, quant.; e) KOAc, DMF, reflux, then KOH, MeOH, reflux, 83%.

to provide the 24S-hydroxy product in 85% yield and >99.9% *de*<sup>15</sup> The high yield and *de* may be owing to the special structure of steroidal aldehyde **3** or the novel ligand or both. Acetylation of **4** with acetic anhydride in the presence of pyridine gave the acetate **5** in 99% yield, then removal of 3 $\alpha$ , 6 $\alpha$ -dihydroxy protective groups by refluxing with PPTS in <sup>t</sup>BuOH afforded the 3 $\alpha$ , 6 $\alpha$ -dihydroxy cholesterol (**6**) in 88% yield. Ditosylation of **6** with *p*-toluene sulfonyl chloride in pyridine at 0 °C provided quantitatively the ditosylate **7** which was treated with potassium acetate in DMF-water at 105 °C<sup>13</sup> followed by hydrolysis of the ester with potassium hydroxide in methanol to give the target molecule cerebrosterol (**1**) in 83% yield.<sup>16</sup>

The advantages of this synthesis of cerebrosterol are the easy availability of starting material (Me-HDCA) and a short route with high diastereoselectivity (>99.9% *de*).

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## References and notes

- Ercoli, A.; DeRuggieri, P. *Gazz. Chim. Ital.* **1953**, *83*, 78.
- (a) Lütjohann, D.; Breuer, O.; Ahlborg, G.; Nennesmo, I.; Siden, A.; Diczfalusy, U.; Bjorkhem, I. *Proc. Natl. Acad. Sci. U. S. A.* **1996**, *93*, 9799.  
(b) Bjorkhem, I.; Lütjohann, D.; Breuer, O.; Sakinis, A.; Wennmalm, A. *J. Biol. Chem.* **1997**, *272*, 30178.  
(c) Zhang, J.; Akwa, Y.; El-Etr, M.; Baulieu, E.-E.; Sjovall, J. *Biochem. J.* **1997**, *322*, 175.  
(d) Bjorkhem, I.; Lütjohann, D.; Diczfalusy, U.; Stähle, L.; Ahlborg, G.; Wahren, J. *J. Lipid Res.* **1998**, *39*, 1594.  
(e) Kolsch, H.; Lütjohann, D.; Tulke, A.; Bjorkhem, I.; Rao, M. L. *Brain Res.* **1999**, *818*, 171.  
(f) Bretillon, L.; Lütjohann, D.; Stähle, L.; Widhe, T.; Bindl, L.; Eggertsen, G.; Diczfalusy, U.; Bjorkhem, I. *J. Lipid Res.* **2000**, *41*, 840.
- Lütjohann, D.; Papassotiropoulos, A.; Bjorkhem, I.; Locatelli, S.; Bagli, M.; Oehring, R. D.; Schlegel, U.; Jessen, F.; Rao, M. L.; Von Bergmann, K.; Heun, R. *J. Lipids Res.* **2000**, *41*, 195.
- (a) Lehmann, J. M.; Kliewer, S. A.; Moore, L. B.; Smith-Oliver, T. A.; Oliver, B. B.; Su, J. L.; Sundseth, S. S.; Winegar, D. A.; Blanchard, D. E.; Spencer, T. A.; Willson, T. M. *J. Biol. Chem.* **1997**, *272*, 3137.  
(b) Peet, D. J.; Turley, S. D.; Ma, W.; Janowski, B. A.; Lobaccaro, J.-M. A.; Hammer, R. E.; Mangelsdorf, D. J. *Cell* **1998**, *93*, 693.
- (a) Schultz, J. R.; Tu, H.; Luk, A.; Repa, J. C.; Medina, J. C.; Li, L.; Schwendner, S.; Wang, S.; Thoolen, M.; Mangelsdorf, D. J.; Lustig, K. D.; Shan, B. *Genes Dev.* **2000**, *14*, 2831;  
(b) Repa, J. J.; Liang, G.; Ou, J.; Bashmakov, Y.; Lobaccaro, J. M. A.; Shimomura, I.; Shan, B.; Brown, M. S.; Goldstein, J. L.; Mangelsdorf, D. J. *Genes Dev.* **2000**, *14*, 2819.
- (a) Repa, J. J.; Turley, S. D.; Lobaccaro, J.-M. A.; Medina, J.; Li, L.; Lustig, K.; Shan, B.; Heyman, R. A.; Dietschy, J. M.; Mangelsdorf, D. J. *Science* **2000**, *289*, 1524;  
(b) Costet, P.; Luo, Y.; Wang, N.; Tall, A. R. *J. Biol. Chem.* **2000**, *275*, 28240;  
(c) Venkateswaran, A.; Repa, J. J.; Lobaccaro, J.-M. A.; Bronson, A.; Mangelsdorf, D. J.; Edwards, P. A. *J. Biol. Chem.* **2000**, *275*, 14700;  
(d) Berge, K. E.; Tian, H.; Graf, G. A.; Yu, L.; Grishin, N. V.; Schultz, J.; Kwiterovich, P.; Shan, B.; Barnes, R.; Hobbs, H. H. *Science* **2000**, *290*, 1771.
- Laffitte, B. A.; Repa, J. J.; Joseph, S. B.; Wilpitz, D. C.; Kast, H. R.; Mangelsdorf, D. J.; Tontonoz, P. *Proc. Natl. Acad. Sci. U. S. A.* **2001**, *98*, 507.
- Luo, Y.; Tall, A. R. *J. Clin. Invest.* **2000**, *105*, 513;
- Zhang, Y.; Repa, J. J.; Gauthier, K.; Mangelsdorf, D. J. *J. Biol. Chem.* **2001**, *276*, 43018.
- (a) Ercoli, A.; DeRuggieri, P. *J. Am. Chem. Soc.* **1953**, *75*, 3284.  
(b) Ikekawa, N.; Morisaki, M.; Koizumi, N.; Sawamura, M.; Tanaka, Y.; Deluca, H. F. *Biochem. Biophys. Res. Commun.* **1975**, *62*, 485.
- (a) Koch, P.; Nakatani, Y.; Liu, B.; Ourisson, G. *Bull. Soc. Chim. Fr.* **1983**, *II*, 189.  
(b) Moriarty, R. M.; Enache, L. A.; Kinny, W. A.; Allen, C. S.; Canary, J. W.; Tuladhar, S. M.; Guo, L. *Tetrahedron Lett.* **1995**, *36*, 5139.  
(c) Koizumi, N.; Ishiguro, M.; Yasuda, M.; Ikekawa, N. *J. Chem. Soc., Perkin Trans. 1* **1983**, 1401.  
(d) Okamoto, M.; Tabe, M.; Fujii, T.; Tanaka, T. *Tetrahedron: Asymmetry* **1995**, *6*, 767.  
(e) Corey, E. J.; Grogan, M. J. *Tetrahedron Lett.* **1998**, *39*, 9351.  
(f) Zhou, X. D.; Zhou, W. S. *Tetrahedron* **2001**, *57*, 8291.
- (a) Prasad, V. K.; Ponticorvo, L.; Lieberman, S. J.

- Steroid Biochem.* **1984**, *21*, 733.
- (b) Fukumoto, K.; Suzuki, K.; Nemoto, H.; Kametani, T.; Fukuyama, H. *Tetrahedron* **1982**, *38*, 3704.
- 13 Bharucha, K. R.; Buckley, G. C.; Cross, C. K.; Rubin, L. J.; Ziegler, P. *Can. J. Chem.* **1956**, *34*, 982.
- 14 Yang, W. K.; Cho, B. T. *Tetrahedron: Asymmetry* **2000**, *11*, 2947.
- 15 (a) Compound **4**:  $[\alpha]_D^{20} + 4.8$  (*c* 0.965, CHCl<sub>3</sub>); MS (70 eV) *m/z* (%): 444 (8.9), 384 (53.8), 45 (100.0); IR (KBr)  $\nu$ : 3503 (OH) cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz)  $\delta$ : 0.64 (s, 3H, 18-CH<sub>3</sub>), 0.89 (d, *J* = 6.8 Hz, 3H, 21-CH<sub>3</sub>), 0.911–0.920 (m, 6H, 19-CH<sub>3</sub>, 26-CH<sub>3</sub>), 0.94 (d, 3H, *J* = 2.1 Hz, 27-CH<sub>3</sub>), 3.30–3.36 (m, 1H, 24-H), 3.37 (s, 3H, OCH<sub>3</sub>), 3.38 (s, 3H, OCH<sub>3</sub>), 3.45–3.59 (m, 1H, 3 $\beta$ -H), 3.90–3.94 (m, 1H, 6 $\beta$ -H), 4.65–4.74 (m, 4H, OCH<sub>2</sub>O  $\times$  2). Anal. calcd for C<sub>31</sub>H<sub>56</sub>O<sub>5</sub>: C 73.18, H 11.09; found C 73.02, H 10.77. The diastereoselective ratio of the product was determined by HPLC analysis on Inersil ODS column (4.6  $\times$  250 mm) with CH<sub>3</sub>CN as eluent. The absolute configuration was identified by comparison with authentic sample.<sup>15b</sup>
- (b) Zhou, X. D. *Ph. D. Dissertation*, Shanghai Institute of Organic Chemistry, Shanghai, **2001** (in Chinese).
- 16 Compound **1**: yield 83%; m.p. 173–174 °C (lit.<sup>10a</sup> m.p. 175–176 °C; lit.<sup>17</sup> m.p. 181–182.5 °C);  $[\alpha]_D^{20} - 48$  (*c* 0.245, CHCl<sub>3</sub>) (lit.<sup>17</sup>  $[\alpha]_D - 48.3$ , CHCl<sub>3</sub>); MS (70 eV) *m/z* (%): 402 (52.4), 384 (38.8), 369 (18.0), 351 (14.7); IR (KBr)  $\nu$ : 3370 (OH) cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz)  $\delta$ : 0.68 (s, 3H, 18-CH<sub>3</sub>), 0.89 (d, *J* = 6.8 Hz, 3H, 21-CH<sub>3</sub>), 1.01 (s, 3H, 19-CH<sub>3</sub>), 3.28–3.35 (m, 1H, 24-H), 3.47–3.58 (m, 1H, 3 $\alpha$ -H), 5.35 (d, *J* = 4.8 Hz, 1H, 6-H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz)  $\delta$ : 140.8 (5-C), 121.8 (6-C), 77.5 (24S-C), 77.3 (24R-C), 71.9 (3-C), 56.8 (14-C), 56.0 (17-C), 50.2 (9-C), 42.5 (13-C), 42.4 (4-C), 39.9 (12-C), 37.3 (1-C), 36.6 (10-C), 36.0 (20-C), 33.2 (25-C), 32.3 (22-C), 32.1 (2-C), 32.0 (8-C), 31.7 (7-C), 30.8 (23-C), 28.3 (16-C), 24.3 (15-C), 21.2 (11-C), 19.5 (19-C), 19.1 (21-C), 18.9 (17-C), 16.8 (26-C), 12.0 (18-C). HREIMS calcd for C<sub>27</sub>H<sub>46</sub>O<sub>2</sub>[M<sup>+</sup>], 402.3498, found 402.3533.
- 17 Saucier, S. E.; Kandutsh, A. A.; Clark, D. S.; Spencer, T. A. *Biochem. Biophys. Acta* **1993**, *1166*, 115.

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